

Quantum-chemical Calculations of the Antioxidant Properties of *trans-p-coumaric* Acid and *trans-sinapinic* Acid

Alicja Urbaniak^{1*}, Marcin Molski¹, Małgorzata Szela²

¹Department of Theoretical Chemistry, Faculty of Chemistry,
A. Mickiewicz University, Grunwaldzka 6, 60-780 Poznań, Poland

²Department of Human Molecular Genetics, Faculty of Biology,
A. Mickiewicz University, Umultowska 89, 61-614 Poznań, Poland

*e-mail: au50380@st.amu.edu.pl

(Received: 11 February 2012; accepted: 6 June 2012; published online: 27 August 2012)

Abstract: *Trans-p-coumaric* and *trans-sinapinic* acids are cinnamic acid's derivatives which show strong antioxidant properties. In this work full optimization of both chemical structures and their radical, cation radical and anionic forms in vacuum and water medium has been performed, and antioxidant descriptors: Bond Dissociation Enthalpy, Adiabatic Ionization Potential, Proton Dissociation Enthalpy, Proton Affinity, Electron Transfer Enthalpy, Gas Phase Acidity, Free Gibbs Energy have been calculated. The Highest Occupied and Lowest Unoccupied Molecular Orbital energies have been employed to determine groups in compounds studied with the highest electron density. All calculations were performed using Gaussian 03W software package at the DFT level of theory (B3LYP hybrid functional) together with 6-311+G(2d,2p) basis set. Strong antioxidant properties of both investigated compounds were determined in this study. Based on the results it may be suggested that *trans-p-coumaric* and *trans-sinapinic* acids react according to the Hydrogen Atom Transfer mechanism in vacuum and according to Single-Electron Transfer followed by the Proton Transfer mechanism in water medium. Moreover, in both compounds studied the O-H bond is a preferred place of free radical attack.

Key words: *trans-p-coumaric* acid, *trans-sinapinic* acid, antioxidant properties, DFT, C-PCM model, BDE, AIP, PDE, PA, ETE, HOMO and LUMO energy

I. INTRODUCTION

Free radicals are very reactive chemical species with an unpaired electron. Because of their reactivity lipids, proteins and DNA can be damaged by the radicals action. In consequence, they are responsible for many diseases, such as cancer [1-4], cardiovascular disorders [5-8], atherosclerosis [9-12], asthma, arthritis, neurodegenerative disorders: Alzheimer's [13-17], Parkinson's diseases and dementia [18]. Antioxidants are compounds of natural and synthetic origin with the capacity to scavenge free radicals, hence they have been studied intensively.

The *trans-p-coumaric* acid (3-(4-hydroxyphenyl)prop-2-enoic acid, Fig. 2.) as well as *trans-sinapinic* acid (3-(4-hydroxy-3,5-dimethoxyphenyl)prop-2-enoic acid Fig. 3) are cinnamic acid's ((*E*)-3-phenylprop-2-enoic acid, Fig. 1) derivatives. Both substances occur in nature: the *trans-p-*

coumaric acid is present in plants (for example, in bamboo leaves [19]), microorganisms, animals [20], and is synthesized in the human liver [21], whereas the *trans-sinapinic* acid is present in edible plants and fruits [22-25] (broccoli, leafy brassicas and citrus juices). Antioxidant properties of the *trans-p-coumaric* acid were studied experimentally and theoretically [21, 26]. Because of its radical scavenging activity the *trans-p-coumaric* acid can reduce serum cholesterol level and decrease lipid peroxidation [21]. Moreover, together with caffeic and ferulic acids it can promote the excretion of natural sterols which leads to decreased absorption of dietary cholesterol [27]. The *trans-sinapinic* acid is a strong antioxidant [28-30], which is endowed also with anxiolytic [31] and anti-inflammatory properties [32]. The ability of the sinapinic acid to inhibit peroxynitrite-mediated oxidation related with its antioxidant properties has been confirmed by *in*

in vitro experiments [33-34]. In comparison to other phenolic antioxidants, the sinapinic acid has stronger antioxidant properties than the ferulic acid, syringic acid and p-coumaric acid but a little bit lower than the curcumin and chlorogenic acid and much lower than caffeic and gallic acids [35-37]. For instance, in the DPPH (2,2-diphenyl-1-picrylhydrazyl) analysis the antioxidant properties of compounds change as: gallic acid > caffeic acid ~ ascorbic acid ~ Trolox > sinapinic acid > isoeugenol [37]. ABTS (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) assay has shown that the sinapinic acid is a weaker antioxidant than the gallic acid but stronger than the rest of tested compounds [37]. CBA (Crocin Bleaching Assay) test has shown that the sinapinic acid is the strongest antioxidant of all investigated compounds [37]. In ORAC assay

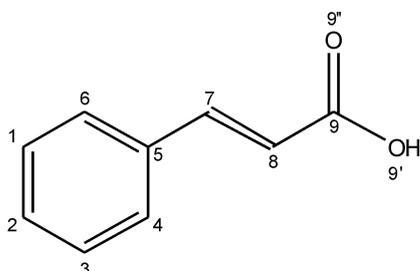


Fig. 1. Molecular structure of *trans*-cinnamic acid

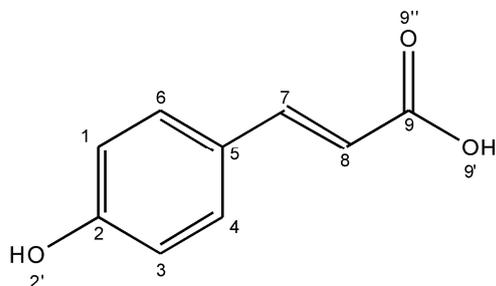


Fig. 2. Molecular structure of *trans*-*p*-coumaric acid

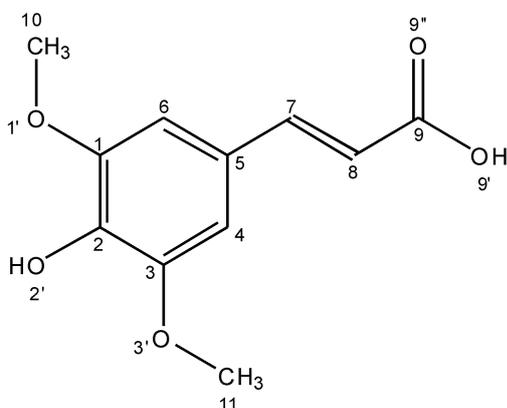
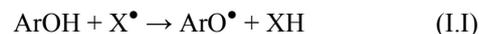


Fig. 3. Molecular structure of *trans*-sinapinic acid

(Oxygen Radical Absorbance Capacity) only caffeic and isoeugenol have stronger antioxidant abilities than the sinapinic acid [37].

There are five known antioxidant mechanisms which describe antioxidant reactions [38-47]:

1) HAT (*Hydrogen atom transfer*) mechanism:



According to this mechanism phenolic antioxidant reacts directly with a free radical which is neutralized, and a radical form of phenolic antioxidant appears. A numerical parameter associated with this mechanism is BDE (*Bond Dissociation Enthalpy*). The lower BDE parameter characterizes better antioxidant property.

2) SET (*Single electron transfer*) mechanism:



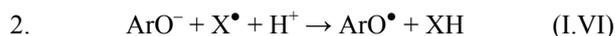
A numerical parameter related with the SET mechanism is AIP (*Adiabatic Ionization Potential*).

3) SET-PT (*Single-electron transfer followed by proton transfer*):



This mechanism is a two-step reaction. In the first step a phenolic antioxidant molecule reacts with the free radical, and a cationic radical form of the phenolic antioxidant and an anionic form of the radical appear. This reaction is a thermodynamically significant step of this two-step mechanism. In the second step the cationic radical form of the phenolic antioxidant decomposes into a phenolic radical and proton. A numerical parameter related with the SET-PT mechanism is AIP (*Adiabatic Ionization Potential*) for the first step and PDE (*Proton Dissociation Enthalpy*) for the second step.

4) SPLET (*Sequential proton loss electron transfer*):



This mechanism also consists of a two-step reaction. In the first step the phenolic antioxidant dissociates into an anionic form and proton, and then ions created in the first reaction react with the free radical. In this reaction a radical form of the phenolic antioxidant and a neutral molecule appear. A numerical parameter related with this mechanism is for the first reaction step: PA (*Proton affinity*) and for the second step: ETE (*Electron Transfer Enthalpy*). Coexistence of the presented mechanisms is shown in Fig. 4.

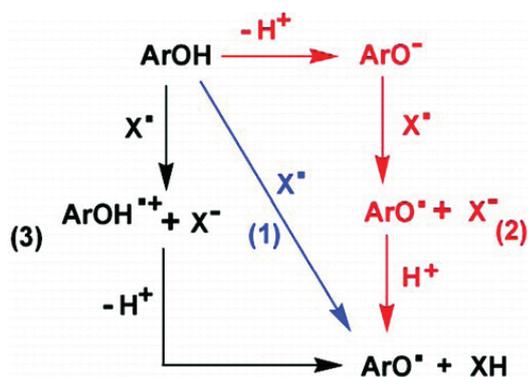


Fig. 4. Phenolic antioxidants-mechanisms of action: (1) HAT, (2) SPLET, (3) SET-PT. ArOH - phenolic antioxidant, X^\bullet - free radical. From [48]

The other mechanism used to describe antioxidant properties is TMC (*Transition Metals Chelation*) [45]. Metals in their low oxidation state may generate free radicals according to the Fenton reaction:



In the TMC scheme each molecule which may dissociate has the ability to chelate heavy metals. Especially anions of polyphenols have the significant abilities to chelate heavy metals. Since chelation of metals often occurs due to deprotonated hydroxyls in the polyphenols, the ability of a molecule to produce the proton is taken into consideration. The numerical parameter related with this mechanism is *gas phase acidity* – it is identified with the vacuum enthalpy of the compound $\Delta H_{\text{acidity}}$. For the calculations in solvents the free Gibbs energy $\Delta G_{\text{acidity}}$ is calculated.

Other important descriptors of the antioxidant properties are distributions of HOMO and LUMO and spin density – SD of molecules. The lower HOMO energy is responsible for the poorer abilities of a molecule to donate a proton. Hence, the HOMO distribution reveals which chemical groups in a molecule are easily attacked by free radicals. The SD parameter characterizes the distribution of non-paired electron and the stability of a radical form of the molecule. From the difference between LUMO and HOMO energy we can conclude about chemical activity of the molecule. The lower $\Delta E(\text{LUMO} - \text{HOMO})$ is connected with lower activity of the molecule [49].

The main aim of this work was to optimize structures of all the studied compounds: neutral, radical, cationic radical and anionic forms to explain the structure-antioxidant relationship. We had also been concerned with the calculation of antioxidant descriptors: BDE, AIP, PDE,

PA, ETE, $\Delta H_{\text{acidity}}$, $\Delta G_{\text{acidity}}$ for *trans-p-coumaric* and *trans-sinapinic* acids. The next step was the determination of the preferred mechanism of antioxidation and calculation of HOMO and LUMO energies and transition energy between those states. All calculations were performed in vacuum and water medium.

II. COMPUTATIONS

All quantum-chemical calculations were performed using the Gaussian 03W [50] software package. Geometry of *trans-p-coumaric* and *trans-sinapinic* acids in their ground state were fully optimized in vacuum and water environment employing the DFT method with restricted B3LYP hybrid functional [51-53] together with 6-311+G(2d,2p) basis set. However, for the optimization of the geometry of the radicals and cation radicals the unrestricted B3LYP/6-311+G(2d,2p) level of theory was applied. The optimization of the geometry of mono- and dianions was carried out with the rB3LYP/6-311+G(2d,2p). To decide which antioxidant mechanism is preferred for these two compounds the numerical descriptors: BDE, AIP, PDE, PA, ETE, $\Delta H_{\text{acidity}}$, $\Delta G_{\text{acidity}}$ defined below have been calculated.

$$\text{BDE} = H_{\text{ArO}^\bullet} + H_{\text{H}^\bullet} - H_{\text{ArOH}} \quad (\text{II.I})$$

in which H_{ArO^\bullet} is the enthalpy of the radical, H_{H^\bullet} is the enthalpy of the H atom, H_{ArOH} is the enthalpy of the compound.

$$\text{AIP} = H_{\text{ArOH}^{\bullet+}} - H_{\text{ArOH}} \quad (\text{II.II})$$

here $H_{\text{ArOH}^{\bullet+}}$ is the enthalpy of cationic radical, H_{ArOH} is the enthalpy of the compound.

$$\text{PDE} = H_{\text{ArO}^\bullet} + H_{\text{H}^+} - H_{\text{ArOH}^{\bullet+}} \quad (\text{II.III})$$

in which H_{ArO^\bullet} is the enthalpy of the radical, H_{H^+} is the enthalpy of the proton, $H_{\text{ArOH}^{\bullet+}}$ is the enthalpy of cationic radical.

$$\text{PA} = H_{\text{ArO}^-} + H_{\text{H}^+} - H_{\text{ArOH}} \quad (\text{II.IV})$$

here H_{ArO^-} is the enthalpy of the anion, H_{H^+} is the enthalpy of the proton, H_{ArOH} is the enthalpy of the compound.

$$\text{ETE} = H_{\text{ArO}^\bullet} - H_{\text{ArO}^-} \quad (\text{II.V})$$

here H_{ArO^\bullet} is the enthalpy of the radical, H_{ArO^-} is the enthalpy of the anion.

$$\Delta H_{\text{acidity}} = H_{\text{ArO}^-} - H_{\text{ArOH}} \quad (\text{II.VI})$$

in which H_{ArO^-} is the enthalpy of the anion, H_{ArOH} is the enthalpy of the compound.

$$\Delta G_{\text{acidity}} = G_{\text{ArO}^-} - G_{\text{ArOH}} \quad (\text{II.VII})$$

here G_{ArO^-} is the Gibbs free energy of the anion, G_{ArOH} is the Gibbs free energy of the compound. To calculate the energy difference ΔE between LUMO and HOMO energies the following formula has been used:

$$\Delta E = E_{\text{LUMO}} - E_{\text{HOMO}} \quad (\text{II.VIII})$$

All calculations carried out in water environment were performed with the C-PCM solvation model (*conductor-like polarizable continuum model*) [54]. The enthalpy values: $H(\text{H}^\bullet)_{\text{vacuum}} = -0.49764$ Ha (hartree) [55], $\Delta_{\text{hydr}}H(\text{H}^\bullet) = -0.00152$ Ha [56-57], $H(\text{H}^+)_{\text{vacuum}} = 0.00236$ Ha [58], $\Delta_{\text{hydr}}H(\text{H}^+) = -0.41516$ Ha [59] were employed in calculations. Parameters describing the electron-donating

abilities of *trans-p*-coumaric and *trans*-sinapinic acids (HOMO distribution and the SD distribution) were calculated in vacuum and in water environment at the B3LYP/6-311+G(2d,2p) level of the theory.

III. RESULTS

III. 1. Geometries

Geometry and the total molecular energy optimizations have been carried out in the vacuum without symmetry constraints in the ground state. Through the scanning of dihedral angles $\alpha = \text{H}_8\text{-C}_8\text{-C}_9\text{-O}_9$ and $\theta = \text{C}_4\text{-C}_5\text{-C}_7\text{-H}_7$, the

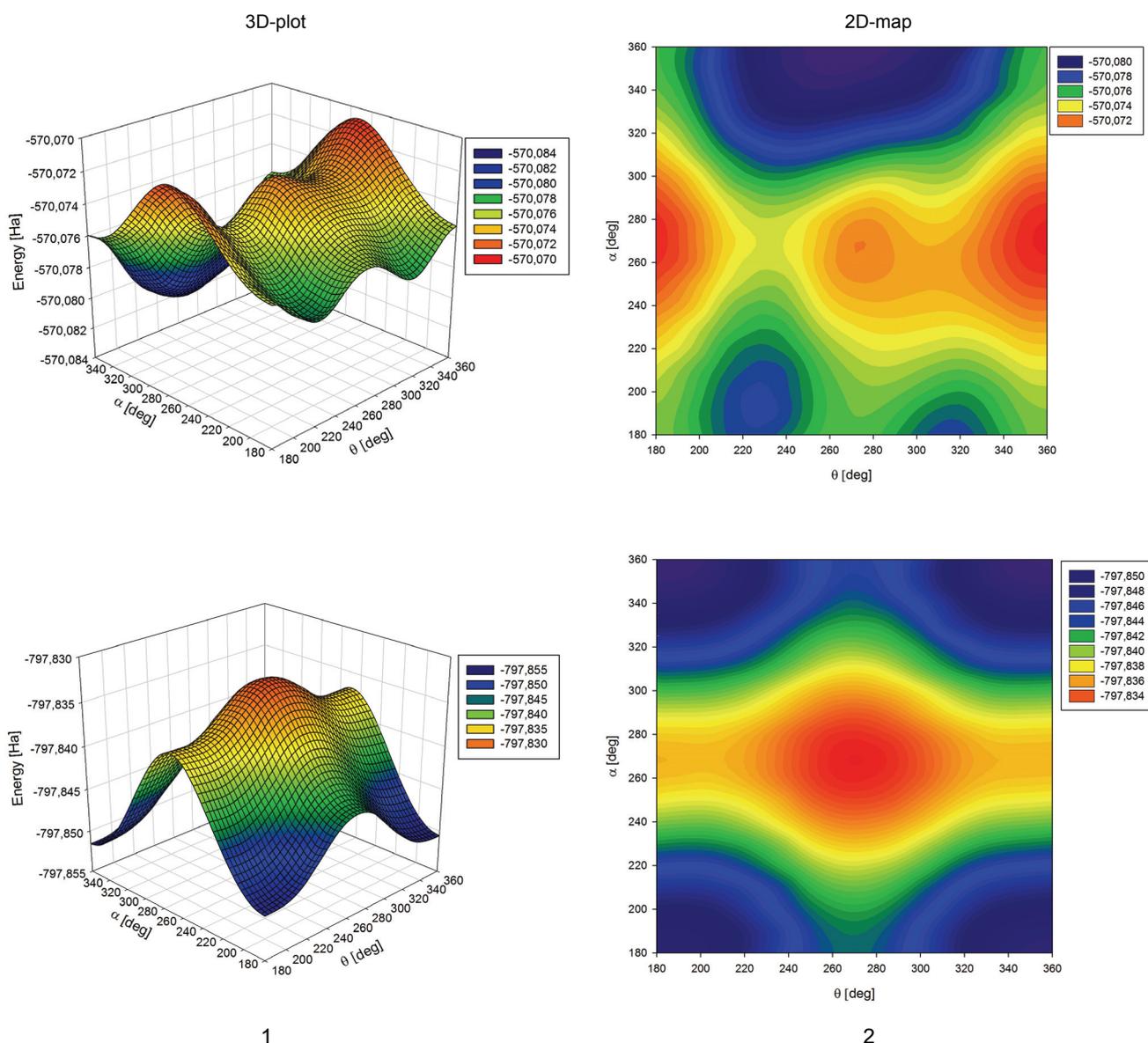


Fig. 5. 3D-plots and 2D-maps of PES for *trans-p*-coumaric acid (1) and *trans*-sinapinic acid (2) in vacuum

Potential Energy Surface profiles (3D-PES) were constructed. PES scans were calculated employing the HF method together with a 6-31G(d,p) basis set (Fig. 5) Dihedral angles were scanned in the range: $180^\circ \leq \alpha \leq 360^\circ$, $180^\circ \leq \theta \leq 360^\circ$. The angles α , θ describe the rotation around the C-C single bonds, as follows: C₈-C₉, C₅-C₇. From the most stable conformations with partial optimization on 3D-PES, the HF/6-31G(d,p) geometry optimization was performed to confirm the positions of the minima. Afterwards, the most stable structures obtained from the energy profiles were fully optimized. Geometry and energy optimization for all studied compounds (Fig. S.1. in Supplement) was made for isolated molecules in vacuum employing the DFT/rB3LYP method together with 6-311+G(2d,2p) basis set. Finally, for each fully optimized structure, the frequency analysis at rB3LYP/6-311+G(2d,2p) was performed to verify that the structures corresponded to stationary points on the PES. Thus the most reliable structures of *trans-p-coumaric* and *trans-sinapinic* acids in their absolute energy minimum have been determined. The same methodology was applied for calculations in water medium using the C-PCM solvation model. In all investigated structures the hydrogen atom from the hydroxyl group is planar with respect to the surface of the molecule. All studied geometries are planar in vacuum and in water medium. The planarity of the geometry influences the antioxidant activity because it causes delocalisation of the lone electron pair through the whole structure: the phenyl ring, vinyl bond and carboxyl group.

III. 2. Descriptors of antioxidant properties

BDE is a numerical parameter related with the HAT mechanism. It characterizes the stability of the bond 2'-O-H in the hydroxyl group. Calculated values of BDE are presented in Table 1. They reveal that the stability of the O-H bond in *trans-p-coumaric* acid is similar in vacuum and in polar medium. The stability of this bond is slightly lower in the *trans-sinapinic* acid than in the *trans-p-coumaric* acid.

Table 1. BDE values [kcal/mol] calculated on B3LYP/6-311+G(2d,2p) level of theory

Bond	BDE	
	Vacuum	Water
<i>trans-p-coumaric</i> acid 2'-O-H	83	85
<i>trans-sinapinic</i> acid 2'-O-H	78	78

The AIP parameter is related to the SET-PT mechanism. It describes the process of electron donation by the antioxidant. Molecules with low AIP values are more susceptible to ionization and have stronger antioxidant properties. The values of AIP have been presented in Table 2. For the *trans-p-coumaric* acid the AIP values are lower in water medium than in vacuum. This fact is related with electrostatic interaction of water medium with cation radical forms of the compounds. The AIP value for the *trans-sinapinic* acid is lower in vacuum than for *trans-p-coumaric* acids. Generally huge AIP values indicate that SET-PT is not the preferred antioxidant mechanism for these two molecules.

Table 2. AIP and PDE values [kcal/mol] calculated on B3LYP/6-311+G(2d,2p) level of theory

Compound	AIP		PDE	
	Vacuum	Water	Vacuum	Water
<i>trans-p-coumaric</i> acid	182	135	215	4
<i>trans-sinapinic</i> acid	172	104	220	2

The PDE parameter characterizes the second step of the SET-PT mechanism. PDE values are also presented in Table 2. The high values of this parameter in vacuum confirm that this mechanism is not preferred. The low values of the PDE parameter for the *trans-p-coumaric* acid in water indicates that this step is energetically favored for this compound in polar medium.

PA and ETE parameters are related with the SPLET mechanism (Table 3). The PA values for both investigated compounds are high in vacuum and small in polar medium. It favors the SPLET mechanism in polar medium for *trans-p-coumaric* and *trans-sinapinic* acids. The second parameter ETE is smaller for both compounds in vacuum than in water medium.

$\Delta H_{\text{acidity}}$ and $\Delta G_{\text{acidity}}$ are parameters related with the TMC mechanism. Values of $\Delta H_{\text{acidity}}$ are comparable for

Table 3. PA and ETE values [kcal/mol] calculated on B3LYP/6-311+G(2d,2p) level of theory

Bond	PA		ETE	
	Vacuum	Water	Vacuum	Water
<i>trans-p-coumaric</i> acid 2'-O-H	328	31	69	108
<i>trans-sinapinic</i> acid 2'-O-H	330	30	62	103

both studied molecules. The huge values of this parameter indicate that this mechanism is not preferred for those two compounds in vacuum. $\Delta G_{\text{acidity}}$ is connected with calculations performed in solvents. The huge value of $\Delta G_{\text{acidity}}$ demonstrates that the TMC mechanism is also not preferred in water medium.

Table 4. $\Delta H_{\text{acidity}}$ and $\Delta G_{\text{acidity}}$ values [kcal/mol] calculated on B3LYP/6-311+G(2d,2p) level of theory

Bond	$\Delta H_{\text{acidity}}$ (Vacuum)	$\Delta G_{\text{acidity}}$ (Water)
<i>trans-p</i> -coumaric acid 2'-O-H	326	291
<i>trans</i> -sinapinic acid 2'-O-H	328	290

On the basis of the performed computations we predict that for *trans-p*-coumaric and *trans*-sinapinic acids the most preferred mechanism in vacuum is HAT, and in water solution – SPLET.

III. 3. HOMO - LUMO distribution and SD

The energies of HOMO and LUMO are not antioxidant descriptors but can be connected to the antioxidant activity of molecules. Higher HOMO energies indicate better electron-donating properties of a molecule. The E_{HOMO} value of *trans-p*-coumaric acid is slightly lower than for the *trans*-sinapinic acid in vacuum as well as in water environment (Table 5). The electronic density of these orbitals in vacuum and in water medium is concentrated on

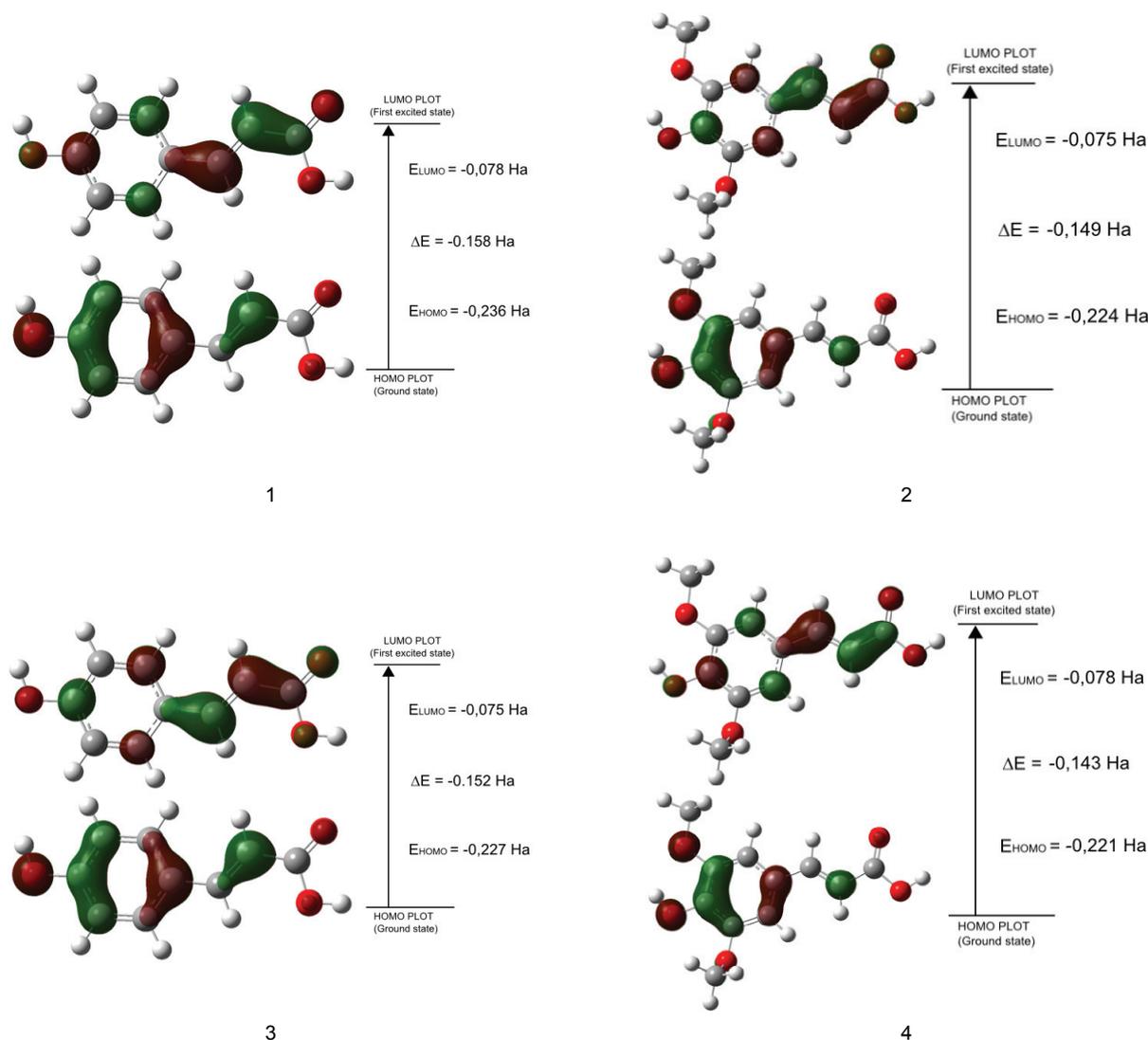


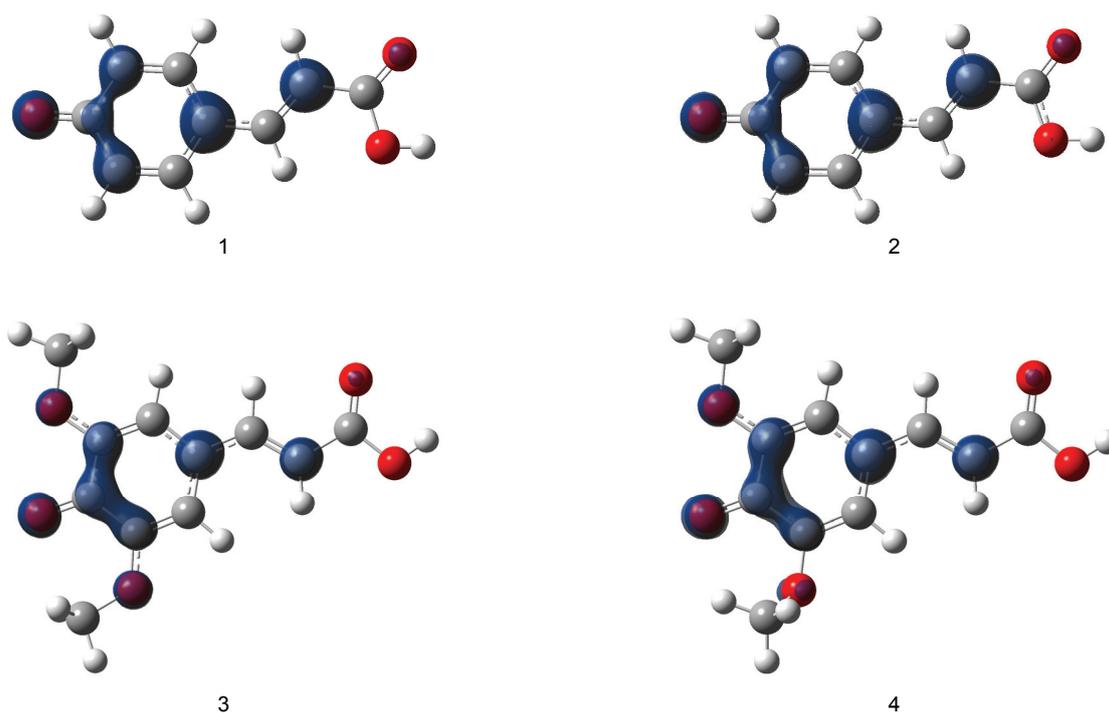
Fig. 6. The atomic orbital compositions of the frontier molecular orbital for *trans-p*-coumaric acid in vacuum (1) and in water medium (2) and for *trans*-sinapinic acid in vacuum (3) and in water medium (4)

Table 5. The HOMO energy (E_{HOMO}) of *trans-p-coumaric acid* and *trans-sinapinic acid* obtained on rB3LYP/6-311G+(2d,2p) level in vacuum and in water medium

Compound	Vacuum		Water	
	E_{HOMO} [Ha]	E_{HOMO} [eV]	E_{HOMO} [Ha]	E_{HOMO} [eV]
<i>trans-p-coumaric acid</i>	-0.236	-6.4	-0.227	-6.2
<i>trans-sinapinic acid</i>	-0.224	-6.1	-0.221	-6.0

Table 6. The LUMO energy (E_{LUMO}) of *trans-p-coumaric acid* and *trans-sinapinic acid* obtained on rB3LYP/6-311G+(2d,2p) level in vacuum and in water medium

Compound	Vacuum		Water	
	E_{LUMO} [Ha]	E_{LUMO} [eV]	E_{LUMO} [Ha]	E_{LUMO} [eV]
<i>trans-p-coumaric acid</i>	-0.078	-2.1	-0.075	-2.0
<i>trans-sinapinic acid</i>	-0.075	-2.0	-0.078	-2.1

Fig. 7. The uB3LYP/6-311+G(2d,2p) SD distribution for 2'-O-radical of *trans-p-coumaric acid* in vacuum (1) and in water medium (2) and 2'-O-radical of *trans-sinapinic acid* in vacuum (3) and in water medium (4). The calculations were accomplished with isovalue 0.004

the oxygen from the hydroxyl group, phenyl ring and vinyl bond in the *trans-p-coumaric acid* (Fig. 6). In case of the *trans-sinapinic acid* the electron density concentrates on the oxygen from the hydroxyl group and methoxy groups, phenyl ring and vinyl bond (Fig. 6). It can be concluded that the O-H bond is the most probable place of a free-radical reaction in the whole molecule in the studied compounds. The energy difference between LUMO and HOMO orbital energies determines the chemical reactivity. This difference shows how easy the transition is from the ground to the excited state (Table 7 and Fig. 6), and how it is related to the stability of radical forms of molecules. Analysis of spin density in 2'-O-radicals of molecules studied shows huge spin density concentration on 2'-O

atoms and carbon atoms in a vinyl bond (Fig. 7 and Fig. 8). Groups with high spin density concentration are sensitive to a free-radical attack.

Table 7. ΔE ($E_{\text{LUMO}} - E_{\text{HOMO}}$) calculated using rB3LYP/6-311G+(2d,2p) for *trans-p-coumaric acid* and ΔE for *trans-sinapinic acid* calculated in vacuum and in water medium

Compound	Vacuum		Water	
	ΔE [Ha]	ΔE [eV]	ΔE [Ha]	ΔE [eV]
<i>trans-p-coumaric acid</i>	0.158	4.3	0.152	4.2
<i>trans-sinapinic acid</i>	0.149	4.1	0.143	3.9

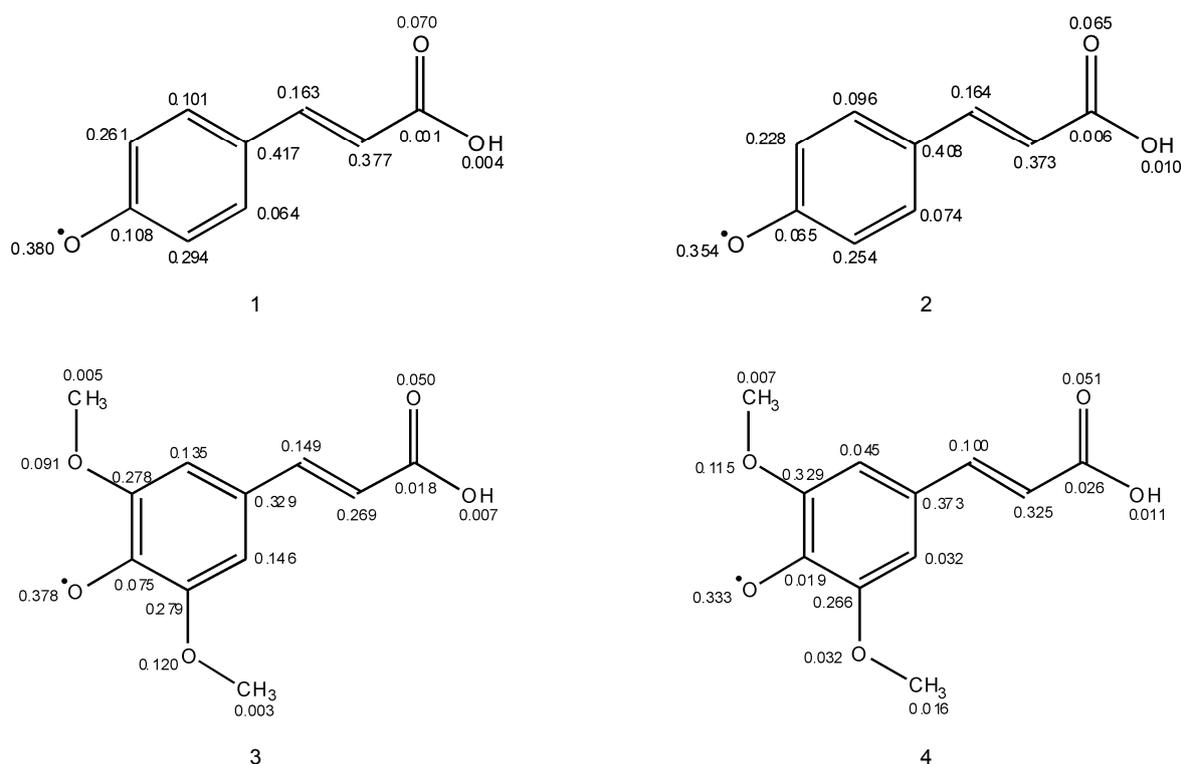


Fig.8. The uB3LYP/6-311+G(2d,2p) atomic SD values for 2'-O-radical of *trans-p*-coumaric acid in vacuum (1) and in water medium (2) and 2'-O-radical of *trans*-sinapinic acid in vacuum (3) and in water medium (4)

IV. CONCLUSIONS

Structure-antioxidant relationships of the *trans-p*-coumaric and *trans*-sinapinic acids have been investigated employing the DFT/B3LYP method together with the 6-311+G(2d,2p) basis set in vacuum and in water medium. Based on the obtained results we conclude that the O-H group is accountable for antioxidant abilities. We also suggest that for both investigated compounds the HAT mechanism is favored in vacuum whereas SPLET is favored in water medium. Quantum chemical calculations confirmed high antioxidant activity of both considered compounds.

Acknowledgements

This study is supported by Poznań Supercomputing and Networking Center (PCSS). The authors are grateful for grants No. 85, "Theoretical investigations of antioxidant activity of cinnamic acid's derivatives" and no. 130 "Theoretical investigations of antioxidant activity of ferulic acid's derivatives".

References

- [1] N.F. Boyd, V. McGuire, *The possible role of lipid peroxidation in breast cancer risk*. Free Radical Biol. Med. 10, 185-190 (1991).
- [2] R.L. Nelson, *Dietary iron and colorectal cancer risk*. Free Radical Biol. Med. 12, 161-168 (1992).
- [3] P. Knekt, A. Reunanen, H. Takkunen, A. Aromaa, M. Heliovaara, T. Hakuunen, *Body iron stores and risk of cancer*. Int. J. Cancer 56, 379-382 (1994).
- [4] G.S. Omenn, G.E. Goodman, M.D. Thornquist, J. Balmes, M.R. Cullen, A. Glass, J.P. Keogh, F.L. Meyskens, B. Valanis, J.H. Williams, S. Barnhart, S. Hammar, *Effects of a combination of beta carotene and vitamin A on lung cancer and cardiovascular disease*. N. Engl. J. Med. 334, 1150-1155 (1996).
- [5] R.A. Riemersma, D.A. Wood, C.C.A. Macityre, R.A. Elton, K.F. Gey, M.F. Oliver, *Risk of angina pectoris and plasma concentrations of vitamins A, C and E and carotene*. Lancet 337, 1-4 (1991).
- [6] J.T. Salonen, K. Nyyssoner, H. Korpela, J. Tuomilehto, R. Seppanen, R. Salonen, *High stored iron levels are associated with excess risk of myocardial infarction in eastern Finnish men*. Circulation 86, 803-811 (1992).
- [7] D.A. Street, G. Comstock, R. Salkeldy, M. Klag, *Serum antioxidants and myocardial infarction. Are low levels of carotenoids and alpha-tocopherol risk factors for myocardial infarction?* Circulation 90, 1154-1161 (1994).
- [8] L.H. Kushi, A.R. Folsom, R.J. Prineas, P. J. Mink, Y. Wu, R. Bostick, *Dietary antioxidant vitamins and death from*

- coronary heart disease in postmenopausal women. *N. Engl. J. Med.* 334, 1156-1162 (1996).
- [9] O.M. Panasenko, T.V. Nova, O.A. Azizova, Y.A. Vladimirov, *Free radical modification of lipoproteins and cholesterol accumulation in cells upon atherosclerosis*. *Free Radical Biol. Med.* 10, 137-148 (1991).
- [10] D. Steinberg, *Antioxidants and Atherosclerosis A Current Assessment*. *Circulation* 84, 1420-1425 (1991).
- [11] D.R. Janero, *Therapeutic potential of vitamin E in pathogenesis of spontaneous atherosclerosis*. *Free Radical Biol. Med.* 11, 129-144 (1991).
- [12] H.N. Hodis, W.J. Mack, L. LaBree, L. Cashin-Hemphill, A. Sevanian, R. Johnson, S. Azen, *Serial coronary angiographic evidence that antioxidant vitamin intake reduces progression of coronary artery atherosclerosis*. *J. Am. Med. Assoc.* 273, 1849-1854 (1995).
- [13] D.A. Butterfield, K. Hensley, M. Harris, M. Mattson, J. Carney, *A model for beta-amyloid aggregation and neurotoxicity based on the free radical generating capacity of the peptide: implications of "molecular shrapnel" for Alzheimer's disease*. *Proc. West Pharmacol. Soc.* 38, 113-120 (1995).
- [14] K. Hensley, J.M. Carney, M.P. Mattson, M. Aksenova, M. Harris, J.F. Wu, R.A. Floyd, D.A. Butterfield, *A model for beta-amyloid aggregation and neurotoxicity based on free radical generation by the peptide: relevance to Alzheimer disease*. *Proc. Natl. Acad. Sci. U. S. A.* 91, 3270-3274 (1994).
- [15] D.A. Butterfield, L. Martin, J. M. Carney, K. Hensley, *A beta (25-35) peptide displays H₂O₂-like reactivity towards aqueous Fe²⁺, nitroxide spin probes, and synaptosomal membrane proteins*. *Life Sci.* 58, 217-228 (1995).
- [16] D.A. Butterfield, *beta-Amyloid-associated free radical oxidative stress and neurotoxicity: implications for Alzheimer's disease*. *Chem. Res. Toxicol.* 10, 495-506 (1997).
- [17] R. Sultana, D.A. Butterfield, *Redox proteomics studies of in vivo amyloid beta-peptide animal models of Alzheimer's disease: Insight into the role of oxidative stress*. *Proteom. Clinic. Appl.* 2, 685-696 (2008).
- [18] C. Rice-Evans, C. Rice-Evans, B. Halliwell, G.G. Lunt, *Free Radicals and Oxidative Stress: Environment, Drugs and Food Additives*. Portland Press, 103-116 (1995).
- [19] Y. Zhang, X. Tie, B. Bao, X. Wu, Y. Zhang, *Metabolism of flavone C-glucosides and p-coumaric acid from antioxidant of bamboo leaves (AOB) in rats*. *B. J. Nutr.* 97, 484-494 (2007).
- [20] F. Borges, F. Roleira, N. Milhazes, L. Santana, E. Uriarte, *Simple coumarins and analogues in medicinal chemistry: occurrence, synthesis and biological activity*, *Curr. Med. Chem.* 12, 887-916 (2005).
- [21] L.Y. Zang, G. Cosma, H. Gardner, X. Shi, V. Castranova, V. Vallyathan, *Effect of antioxidant protection by p-coumaric acid on low-density lipoprotein cholesterol oxidation*. *A. J. Physiol.-Cell Physiol.* 279, C954-C960 (2000).
- [22] K.J. Dabrowski, F.W. Sosulski, *Quantitation of Free and Hydrolyzable Phenolic Acids in Seeds by Capillary Gas-Liquid Chromatography*. *J. Agric. Food Chem.* 32, 123-127 (1984).
- [23] F. Shahidi, M. Naczki, *Phenolics in Food and Nutraceuticals*. CRC Press, Boca Raton, FL (2003).
- [24] U. Thiyam, H. Stöckmann, T.Z. Felde, K. Schwarz, *Antioxidative effect of the main sinapic acid derivatives from rapeseed and mustard oil by-products*. *Eur. J. Lipid Sci. Technol.* 108, 239-248 (2006).
- [25] T. Stevanovic, P.N. Diouf, M.E. Garcia-Perez, *Bioactive polyphenols from healthy diets and forest biomass*. *Curr. Nutr. Food Sci.* 5, 264-295 (2009).
- [26] V.D. Kancheva, L. Saso, P.V. Boranova, A. Khan, M.K. Saroj, M.K. Pandey, S. Malhotra, J.Z. Nechev, S.K. Sharma, A.K. Prasad, M.B. Georgieva, C. Joseph, A.L. DePass, R.C. Rastogi, V.S. Parmar, *Structure-activity relationship of dihydroxy-4-methylcoumarins as powerful antioxidants: Correlation between experimental & theoretical data and synergistic effect*. *Biochem.* 92, 1089-1100 (2010).
- [27] Y.H. Yeh, Y.T. Lee, H.S. Hsieh, D.F. Hwang, *Dietary Caffeic Acid, Ferulic Acid and Coumaric Acid Supplements on Cholesterol Metabolism and Antioxidant Activity in Rats*. *J. Food and Drug Anal.* 17, 123-132 (2009).
- [28] A. Galano, M. Francisco-Marquez, J. Raul Alvarez-Idaboy, *Mechanism and kinetics studies on the antioxidant activity of sinapinic acid*. *Phys. Chem. Chem. Phys.* 13, 11199-11205 (2011).
- [29] A. Gaspar, M. Martins, P. Silva, E. M. Garrido, J. Garrido, O. Firuzi, R. Miri, L. Saso, F. Borges, *Dietary Phenolic Acids and Derivatives. Evaluation of the Antioxidant Activity of Sinapic Acid and Its Alkyl Esters*. *J. Agric. Food Chem.* 58, 11273-11280 (2010).
- [30] E.G. Bakalbassis, A. Chatzopoulou, V.S. Melissas, M. Tsimidou, M. Tsolaki, A. Vafiadis, *Ab initio and density functional theory studies for the explanation of the antioxidant activity of certain phenolic acids*. *Lipids* 36, 181-190 (2001).
- [31] B.H. Yoon, J.W. Jung, J.-J. Lee, Y.-W. Cho, C.-G. Jang, C. Jin, T.H. Oh, J.H. Ryu, *Anxiolytic-like effects of sinapic acid in mice*. *Life Sci.* 81, 234-240 (2007).
- [32] K.J. Yun, D.J. Koh, S.H. Kim, S.J. Park, J.H. Ryu, D.G. Kim, J.Y. Lee, K.T. Lee, *Anti-inflammatory effects of sinapic acid through the suppression of inducible nitric oxide synthase, cyclooxygenase-2, and proinflammatory cytokines expressions via nuclear factor-kappaB inactivation*. *J. Agric. Food Chem.* 56, 10265-10272 (2008).
- [33] T. Niwa, U. Doi, Y. Kato, T. Osawa, *Inhibitory mechanism of sinapinic acid against peroxynitrite-mediated tyrosine nitration of protein in vitro*. *FEBS Lett.* 459, 43-46 (1999).
- [34] Y. Zou, A.R. Kim, J.E. Kim, J.S. Choi, H.Y. Chung, *Peroxy-nitrite scavenging activity of sinapic acid (3,5-dimethoxy-4-hydroxycinnamic acid) isolated from Brassica juncea*, *J. Agric. Food Chem.* 50, 5884-5890 (2002).
- [35] M.E. Cuvelier, H. Richard, C. Berset, *Comparison of the antioxidative activity of some acid-phenols: structure-activity relationship*. *Biosci. Biotechnol. Biochem.* 56, 324-325 (1992).
- [36] O. Firuzi, L. Giansanti, R. Vento, C. Seibert, R. Petrucci, G. Marrosu, R. Agostino, L. Saso, *Hypochlorite scavenging activity of hydroxycinnamic acids evaluated by a rapid microplate method based on the measurement of chloramines*. *J. Pharm. Pharmacol.* 55, 1021-1027 (2003).
- [37] N. Nenadis, O. Lazaridou, M.Z. Tsimidou, *Use of reference compounds in antioxidant activity assessment*. *J. Agric. Food Chem.* 55, 5452-5460 (2007).
- [38] M. Szelag, D. Mikulski, M. Molski, *Quantum – chemical investigation of the structure and the antioxidant properties of α -lipoic acid and its metabolites*. *J. Mol. Mod.*, DOI 10.1007/s00894-011-1306-y (2011).

- [39] M. Leopoldini, N. Russo, M. Toscano, *The molecular basis of working mechanism of natural polyphenolic antioxidants*. Food Chemistry 125, 288-306 (2011).
- [40] M.C. Foti, C. Daquino, C. Geraci, *Electron – transfer reaction of cinnamic acids and their methyl esters with the DPPH radical in alcoholic solutions*. J. Org. Chem. 69, 2309-2314 (2004).
- [41] G. Litwinienko, K.U. Ingold, *Abnormal solvent effects on hydrogen atom abstraction. 2. Resolution of the curcumin antioxidant controversy. The role of sequential proton loss electron transfer*. J. Org. Chem. 69, 5888-5896 (2004).
- [42] M. Musialik, G. Litwinienko, *Scavenging of dpph radicals by vitamin E is accelerated by its partial ionization: the role of sequential proton loss electron transfer*. Org. Lett. 7, 4951-4954 (2005).
- [43] I. Nakanishi, T. Kawashima, K. Ohkubo, H. Kanazawa, K. Inami, M. Mochizuki, K. Fukuhara, H. Okuda, T. Ozawa, S. Itoh, S. Fukuzumi, N. Ikota, *Electron – transfer mechanism in radical scavenging reactions by vitamin E model in a protic medium*. Org. & Biomolecular Chem. 3, 626-629 (2005).
- [44] G. Litwinienko, K.U. Ingold, *Solvent effects on the rates and mechanisms of reaction of phenols with free radicals*. Acc. Chem. Res. 40, 222-230 (2007).
- [45] M. Leopoldini, N. Russo, M. Toscano, *Gas and Liquid Phase Acidity of Natural Antioxidants*. J. Agric. Food Chem. 54, 3078-3085 (2006).
- [46] M. Leopoldini, N. Russo, M. Toscano, *The molecular basis of working mechanism of natural polyphenolic antioxidants*. Food Chem. 125, 288-306 (2011).
- [47] D. Mikulski, M. Szeląg, M. Molski, R. Górniak, *Quantum-chemical study on the antioxidation mechanisms of trans-resveratrol reactions with free radicals in the gas phase, water and ethanol environment*. J. Mol. Structure 951, 37-48 (2010).
- [48] Y.J. Shang, Y.P. Qian, X.D. Liu, F. Dai, X.L. Shang, W.Q. Jia, Q. Liu, J.G. Fang, B. Zhou, *Radical – scavenging activity and mechanism of resveratrol – oriented analogues: influence of the solvent, radical, and substitution*. J. Org. Chem. 74, 5025-5031 (2009).
- [49] S. Sebastian, N. Sundaraganesan, S. Manoharan, *Molecular structure, spectroscopic studies and first-order molecular hyperpolarizabilities of ferulic acid by density functional study*. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 74, 312-323 (2009).
- [50] M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Roob, J.R. Cheeseman, J.A. Montgomery, T. Vreven, K.N. Kudin, J.C. Burant, J.M. Milliam, S.S. Iyengar, J. Tomasi, V. Barone, B. Menucci, M. Cossi, G. Scalmani, N. Rega, G.A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J.E. Knox, H.P. Hratchian, J.B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R.E. Stratmann, O. Yazyev, A.J. Austin, R. Cammi, C. Pomelli, J.W. Ochterski, P.Y. Ayala, K. Morokuma, G.A. Voth, P. Salvador, J.J. Dannenberg, V.G. Zakrzewski, S. Dapprich, A.D. Daniels, M.C. Strain, O. Farkas, D.K. Malick, A.D. Rabuck, K. Raghavachari, J.B. Foresman, J.V. Ortiz, Q. Cui, A.G. Baboul, S. Clifford, J. Ciosłowski, B.B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R.L. Martin, D.J. Fox, T. Keith, M.A. Al-Laham, C.Y. Peng, A. Nanayakkara, M. Challacombe, P.M.W. Gill, B. Johnson, W. Chen, M.W. Wong, C. Gonzalez, J.A. Pople, Gaussian 03, revision C02. Gaussian Inc., Wallingford, CT, 2003.
- [51] K. Hohenberg, W. Kohn, *Inhomogeneous electron gas*. Phys. Review B 136, 864-871 (1964).
- [52] W. Kohn, L.J. Sham, *Self – consistent equations including exchange and correlation effects*. Phys. Review A 140, 1133-1138 (1965).
- [53] C. Lee, W. Yang, R.G. Parr, *Development of the Cole – Salvetti correlation energy formula into a functional of the electron density*. Physical Review B 37, 785-789 (1988).
- [54] M. Cossi, N. Rega, G. Scalmani, V. Barone, *Energies, structures, and electronic properties of molecules in solutions with the C-PCM solvation model*. J. Comp. Chem. 24, 669-681 (2003).
- [55] J.S. Wright, E.R. Johnson, G.A. DiLabio, *Predicting the activity of phenolic antioxidants: theoretical method, analysis of substituent effects, and application to major families of antioxidants*. J. Am. Chem. Soc. 123, 1173-1183 (2001).
- [56] V.D Parker, *Homolytic bond (H-A) dissociation free energies in solution-applications of the standard potential of the (H⁺/H) couple*. J. Am. Chem. Soc. 114, 7458-7462 (1992).
- [57] M.M. Bizarro, B.J.C. Cabral, R.M.B. dos Santos, J.A.M. Simoes, *Substituent effects on the O-H bond dissociation enthalpies in phenolic compounds: agreements and controversies*. Pure Appl. Chem. 71, 1249-1256 (1999).
- [58] E. Klein, J. Rimarcik, V. Lukes, *DFT/B3LYP study of the O-H bond dissociation enthalpies and proton affinities of para- and meta-substituted phenols in water and benzene*. Acta Chim. Slovaca 2, 37-51 (2009).
- [59] P.W. Atkins, *Physical chemistry*. 6th edn. Oxford University Press, Oxford (1998).

SUPPLEMENT

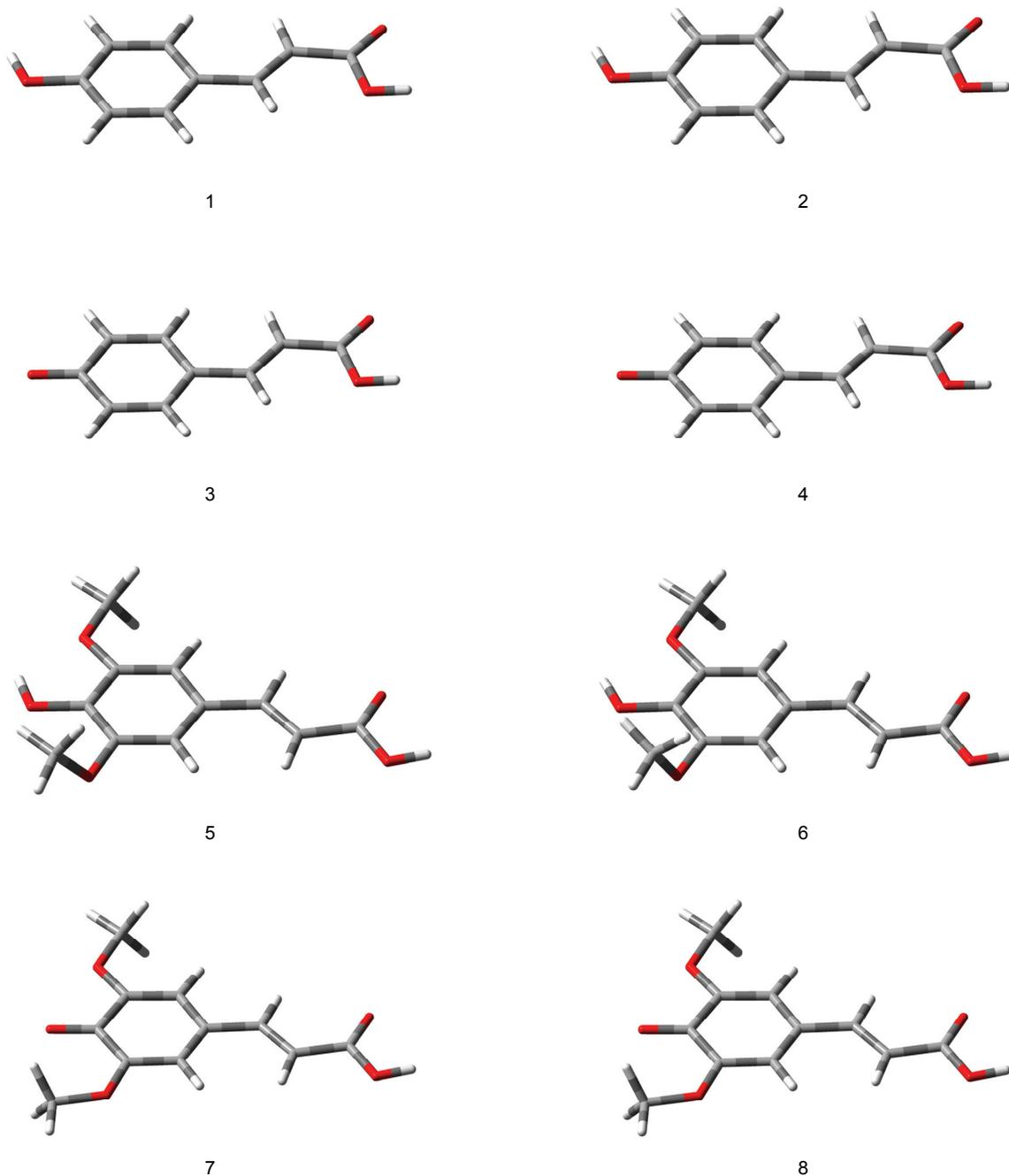


Fig. S.1. The B3LYP/6-311+G(2d,2p) optimized geometries of *trans-p-coumaric acid* in vacuum (1) and in water medium (2) and its 2'-O-radical form in vacuum (3) and in water medium (4) and *trans-sinapinic acid* in vacuum (5) and in water medium (6) and its 2'-O-radical form in vacuum (7) and in water medium (8)



ALICJA URBANIAK, PhD student in chemistry at Adam Mickiewicz University in Poznań. Received Msc degree in chemistry in 2012 at Adam Mickiewicz University in Poznań. Her scientific interests are: quantum chemical calculations of antioxidant properties of substances used in cosmetology.



MARCIN MOLSKI – associate professor at Theoretical Chemistry Department of Adam Mickiewicz University of Poznań. Degrees: PhD chemistry (1992), PhD in Physics (1997), Habilitation (2002). Publications: over 100 scientific papers and 3 books. The research interests concern, among other the following topics: quantum-mechanical determination of chemical, physical and biological properties of bio-active natural substances endowed with antioxidant properties, development of a quasi-quantum theory including macroscopic nonlocal and coherent effects, application of the chaos theory in modeling of biological growth, stimulus-response relationships and cognitive processes in life, behavioral sciences and beyond.



MALGORZATA SZELĄG – PhD student in the Department of Human Molecular Genetics at the Faculty of Biology, Adam Mickiewicz University in Poznan. She received her MSc degree in Biotechnology from UAM in Poznan. Her main areas of interest are: natural antioxidants, QM/MM modeling of interactions between inhibitors and proteins, application of quantum mechanics in drug development.